

人類巨細胞病毒感染與冠狀動脈疾病之相關性

廖永樑¹ 楊義爵¹ 洪鼎鈞¹ 葉慧儀² 陳家鋒³ 廖東南³ 林靖南¹

奇美醫院 ¹病理部 ²內科部 中華醫事學院 ³醫事技術學系

摘 要

人類巨噬細胞病毒(Cytomegalovirus; CMV)可造成無症狀感染，對於免疫不全的個體，更是危及生命的病原。近年有學者證實：冠心病(Coronary artery disease; CAD)屬慢性發炎性疾病，繼而提出慢性感染可能是病因之一，其中又以CMV感染最受矚目。在此篇研究中，我們收集了60位健康正常人和93位冠心病患者共153人，分別偵測其血清中發炎物質濃度、CMV抗體含量、全血中CMV核酸定性及定量。結果顯示：1.冠心病患者體內發炎物質遠高於正常對照組。2.兩組間CMV抗體陽性率並無差別，但若以抗體效價分級，則冠心病組有較高的趨勢。3.CMV PCR陽性率在兩組間並無明顯差異，然而陽性者在兩組之病毒量確有顯著的不同($p < 0.05$)。結論：CMV抗體、CMV nested PCR陽性率與冠心病無關，而在CMV PCR陽性個體，冠心病組血中CMV病毒量較對照組顯著增加。

關鍵詞：人類巨噬細胞病毒 (Cytomegalovirus; CMV)
冠狀動脈疾病 (Coronary artery disease; CAD)
介白素-6 (Interleukin-6; IL-6)
高感度C反應蛋白 (High sensitivity C-reactive protein; Hs-CRP)

引言

由於國人生活型態的改變以及飲食習慣逐漸西化，以致心臟病人數與日俱增。經醫學研究發現，隨著年紀增長，每個人都會出現某種程度的動脈硬化，進而演變成冠狀動脈疾病 (coronary artery disease; CAD)¹⁻³。根據衛生署統計，因心血管疾病而致死的案例，近年來有增加的趨勢，其中又以冠狀動脈疾病最為重要。雖然明確的致病機轉並不清楚，對其高危險致病因子卻已

有深度了解。最常被廣泛討論的危險因子包括：高血壓、有菸癮者、糖尿病、膽固醇過高和缺乏運動者⁴⁻⁶，以及數種發炎因子與生化數值包括：C-reactive protein: CRP、IL-6 homocysteine..等等⁷⁻¹⁰。除此之外，近年來研究發現，受到微生物和病毒感染所引發的發炎反應，如巨細胞病毒 (cytomegalovirus; CMV)等，亦能導致心臟疾病的發生¹¹⁻¹⁴。

巨細胞病毒 (cytomegalovirus; CMV)：屬疱疹病毒科，可感染多種細胞並造成人類相關疾

病，然而大多屬於無症狀感染，無論所造成的疾病種類為何，最後皆轉變成潛伏性感染¹⁵⁻¹⁸。由於CMV 感染具有很高的盛行率，因此易於免疫不全 (immuno-compromised) 的病人身上發病，例如器官移植患者、癌症及 AIDS 患者等。近年研究顯示：CMV 感染在冠狀動脈粥狀化、動脈狹窄等疾病中，扮演一個非常重要的角色，主要是因為原本潛伏在體內的CMV 被活化，使得血管產生發炎反應，進而形成冠狀動脈疾病¹⁹⁻²³。此外，CMV 感染會導致脂質代謝改變，因而使膽固醇累積在動脈中，最後誘發CAD^{24,25}。而氧化態的LDL (low density lipoprotein) 亦能活化上皮細胞、巨噬細胞及平滑肌肉細胞，使得血管發生粥狀化及血栓化效應²⁶⁻²⁹。然而是否引發動脈硬化的關鍵，除了和病毒本身的生物特性有關之外，也與宿主免疫狀態和接受感染的敏感度有密切關係³⁰⁻³²。

雖然CMV 感染與CAD 形成的關聯性已被廣泛的研究，由於該病毒在台灣的盛行率非常高，而且利用血清學方法檢測抗體存在與否，仍會因為個體的變異不同，而產生偽陰性的現象，再加上CMV 難以利用細胞進行培養，根據1999年Douglas 發現，僅60% 的陽性檢體能被培養成功³³⁻³⁵，實在不足以提供臨床診斷與預防醫學之需求。目前臨床上主要以pp65 antigenemia assay 來診斷CMV 感染而造成的疾病，雖然專一性高，但敏感度卻不甚理想³⁶⁻³⁸。因此，有很多研

究CMV 造成的疾病改以PCR 來做診斷，雖然提高了敏感度，但與疾病的專一性卻也降低³⁹⁻⁴²，而且PCR 標定放大基因的種類左右了此方法的敏感度^{40,43,44}。再者，有報告指出：在CMV 抗體陽性及部分陰性的患者血中可檢測出CMV DNA 的存在⁴⁵⁻⁴⁸。然而，亦有相反的研究陳述：在血中以PCR 方式很少將CMV DNA 檢測出來⁴⁹⁻⁵³。本研究將利用目前分子生物學上最新技術—即時聚合酶鏈鎖反應分析儀，(real-time PCR)，偵測不同CAD 病程血液中CMV-DNA 含量，藉以探討病毒量與疾病程或與疾病治療依據之相關。由於real-time PCR 具有快速、高再現性、高敏感性且具有定量功能等優點⁵⁴⁻⁵⁶，冀望能對於診斷心血管疾病或癒後有所助益。

材料及方法

所有的採檢樣本分為二組；對照組60人，包括男性45人，女性15人，均為奇美醫院健康檢查的成人，他們在經過病史理學檢查、一般心電圖和心肌酵素篩檢後，無心絞痛或心肌梗塞之跡象。CAD 組共93人，男性64人，女性29人，為經心導管、運動心電圖或persantin thallium heart scan 證實為冠心病患者。所有採檢樣本的年齡及性別分佈，綜合於(表一)。

IL-6 的測量是採用商業用的成套試劑ELISA (R & D system)，取100 μ l血清和100 μ l的分析稀釋液加入每個well 內 (R & D system ELISA)，

表一：採樣個體分組資料

	Control (n=60)	CAD (n=93)	p value
Age , mean (range)	62.1 (41-83)	64.6 (40-85)	NS
sex , male/female	45/15	64/29	NS
Risk factor , n (%)			
Smoke	12 (21.4)	32 (39.5)	<0.05
Diabetes	10 (16.7)	29 (31.2)	<0.05
Hypertension	21 (35.0)	50 (53.8)	<0.05
Cholesterol , mg/dL	219.8 \pm 37.5 ^a	213.0 \pm 48.6	NS
LDL , mg/dL	143.3 \pm 79.3	125.5 \pm 36.8	NS
TG , mg/dL	117.8 \pm 82.7	196.9 \pm 126.6	<0.001
HDL , mg/dL	56.9 \pm 19.1	44.6 \pm 11.8	<0.001
Bilirubin , mg/dL	0.95 \pm 0.36	0.48 \pm 0.21	<0.001

NS : not significant. p>0.05

^a : mean \pm SD

室溫培育兩個小時。以 Wash Buffer 洗去未結合的物質，再加入抗 IL-6 的多株抗體 200 μ l (conjugated with Horseradish peroxidase)，於室溫培育兩個小時，以 Wash Buffer 洗去未結合的物質，再加入 200 μ l Substrate Solution [H₂O₂ +tetramethylbenzidine mixture (1:1)] 到每一個 well 內，室溫避光培育 20 分鐘，最後加入 50 μ l Stop solution (2N sulfuric acid) 終止呈色反應。以光學比色計測定吸光值 (OD450nm)，求得已知濃度樣品之標準曲線，再定量待測物濃度。每個檢體測量兩次而分析時的變異性小於 10% 。

High sensitivity C-reactive protein (hs-CRP) 實驗方法，利用 Particle-enhanced technology，以 scattered light (Behring BN II nephelometer) 直接偵測抗原抗體複合物濃度而換算出 hs-CRP 含量。

血清中 Bilirubin、uric acid、HDL-C、TG、cholesterol 及 BUN 的測量是利用商業用的成套試劑 (Wake Pure Chemical Industries, Ltd) 以 HITACHI 7070 自動分析儀檢測。

DNA 萃取採用 Puregene DNA purification Kit (Gentra, Minnesota, USA) 抽取全血 (0.3ml) 中的 DNA，最後溶在 0.1ml 純水，保存在 -80 °C。Nested PCR 採用標的基因為 UL122，外部引子分別為 MIE-4 (5'-CCAAGCGGCCTCTGATAAC-CAAGCC-3')，MIE-5 (5'-CAGCACCATCC-TCTCTTCCTCTGG-3') 可放大 435bps 的產物⁵⁷，內部引子依據美商應用生命公司 (ABI) Primer express 軟體設計，針對此產物設計出內部引子，分別為 NS1 (5'-TGAAGGTCTT-TGCCAGTACAT-3')，NAS1 (5'-TGGC-CAAAGTGTAGGCTACAAT-3')，可放大出 129bps 的產物。1st-PCR 試劑調配如下：在 50 μ l 的反應總體積內包含有 30pmol 外部引子、5 μ l DNA 和 1.5U Taq polymerase (Promega, Madison, WI)。2nd-PCR 試劑調配如下：在 50 μ l 的反應總體積內包含有 30pmol 內部引子、1 μ l 1st-PCR 產物和 1.5U Taq polymerase。兩次 PCR 反應條件相同：加熱 95 °C 10 分鐘，三溫度 40 個循環 (95 °C，1 分鐘、55 °C，1 分鐘、72 °C，1 分鐘) 最後在 72 °C 作用 10 分鐘。Nested PCR 產物最終以 7 %

PAGE 進行電泳後，以 ethidium bromide 染色，再 UV 呈色後照相存檔。

Real time PCR 定量分析方面採用 ABI PRISM 7000 偵測系統，以 UL83 為標的基因，引子分別為 pp549s (5'-GTCAGCGTTCGTGTTTCCCA-3')、pp812as (5'-GGGACACAACACCGTAAAGC-3')，探針為 pp770s (5'-FAM-C C C G-CAACCCGCAACCCTTCATG-3'-TAMRA)。標準品備製乃以 pp549s、pp812as 兩引子批出 PCR 產物，轉殖入大腸桿菌內 (TOPO TA Cloning Kit, Invitrogen) 大量繁殖，最後進行質體 DNA 萃取，以 260nm 測定標準品濃度。Real time PCR 試劑調配如下：在 25 μ l 的反應總體積內包含有 12.5pmol 外部引子、5 pmol 探針、5 μ l DNA 和 1U platinum taq。反應條件：加熱 95 °C 10 分鐘，二溫度 45 個循環 (95 °C，15 秒、65 °C，1 分鐘)⁵⁴。

在統計分析方面：在非常態分布的實驗結果，我們使用無母數統計方式。Spearman's rank test 用來評估相關性，而 Chi-Square test 用來評估不連續資料的相關性。以 multiple logistic regression models 調整 CAD 危險因子對生化數值的影響並依序計算出勝率比 (odds ratio, OR) 和 95% 信賴區 (confidence interval, CI)。有意義的顯著值設為 p<0.05。

結果

CAD 的危險因子包括有性別、年齡、肥胖、遺傳、糖尿病、抽煙、高血壓、膽固醇血脂過高、缺乏運動、精神緊張、壓力大。我們隨機選取對照組；健康成人共 60 人：其中男性 45 人，女性 15 人。實驗組；CAD 患者共 93 人：男性佔 64 人，女性 29 人。其中二組的年齡、性別、膽固醇和 LDL 並無顯著的差異，然而二組間之糖尿病、抽煙、高血壓、TG 和 HDL 皆有明顯的差異 (表一)。為避免這些危險因子影響後續的生化數值統計，我們以 multiple logistic regression models 調整這些變數的影響。

發炎分子、CMV 抗體與 PCR 之表現

在 CAD 組中 IL-6、hs-CRP 之濃度較健康對

表二：採樣個體CMV 感染與發炎指標之表現

	Control n (%)	CAD n (%)	p value
Hs-CRP , mg/dL	2.51 ± 0.63 ^a	11.81 ± 2.23	<0.001
IL-6 , pg/ml	1.09 ± 0.40	13.69 ± 4.82	0.002
α CMV IgG , EU/ml	49.2 ± 2.4	56.0 ± 2.3	0.097
Positive>15 EU/ml	59(98.3)	91(97.8)	0.703
Nested PCR	15(25)	29(31.2)	0.409

NS : not significant. $p>0.05$

^a : mean ± SEM

表三：以CMV 抗體效價分成四等分，分析冠心症相對危險性

	Control (n=60)	CAD (n=93)	relative risk (95% CI)	p value
α CMV IgG (EU/ml)				
mean (range)	49.2 (2.8~98.7)	56.0 (9.18~135.4)		
Quartiles distribution , n (%)				
1 (15~30 EU/ml)	8 (13.6%)	6 (7.7%)		
2 (30~60 EU/ml)	35 (59.3%)	49 (53.8%)	1.58 (0.55~4.49)	0.395
3 (60~90 EU/ml)	14 (23.7%)	27 (29.7%)	2.17 (0.69~6.86)	0.187
4 (\geq 90 EU/ml)	2 (3.4%)	9 (9.9%)	5.06 (0.83~30.75)	0.078

p values were calculated by logistic regression analysis.

CI denotes confidence interval.

照組呈現顯著增加，然而anti CMV IgG 抗體效價只有微幅增加且未達顯著意義。同時，兩組間CMV 抗體陽性率皆高達97 % 以上，未有差別。此外，兩組間的CMV DNA PCR 陽性率分別為25 % 與31.2 % ，CAD 組略高於對照組(表二)。

CMV 抗體效價對罹患冠狀動脈疾病的影響

依CMV 抗體效價由低至高分成4 等分(level 1~4)，並分析相對危險性(relative risk)。結果顯示，CAD 組CMV IgG 抗體效價level 2~4 與level 1 相較，其相對危險性分別為1.58 ($p=0.395$)、2.17 ($p=0.187$)、5.06 ($p=0.078$) 有逐漸增加的趨勢(表三)。再以迴歸統計分析CMV 抗體效價、IL-6 、hs-CRP 、PCR 之相關性，結果顯示：在對照組中CMV 抗體效價與IL-6 呈現相關趨勢($r=0.249$, $p=0.055$)。在CAD 組中，僅IL-6 與hs-CRP 呈現高度相關($r=0.634$, $p<0.001$)。但就整體而言：IL-6 與CMV 抗體效價和hs-CRP 皆呈現顯著的相關($r=0.178$, $p=0.028$; $r=0.503$, $p<0.001$) (表四)。

兩組間CMV 病毒量之表現

為檢查病毒量在兩組間是否有差異，我們以定量系統分析血中CMV 病毒量，結果顯示：nested PCR 陰性的檢體，病毒定量依然為陰性(Data not shown)。然而 nested PCR 陽性的檢體，病毒量表現的多寡就有差異。在對照組中，15 個nested PCR 陽性檢體皆測不到CMV 病毒量；而在CAD 組中，29 個nested PCR 陽性檢體就有14 個測得CMV 病毒量($p<0.05$)，範圍由數百至數萬不等(圖一)。

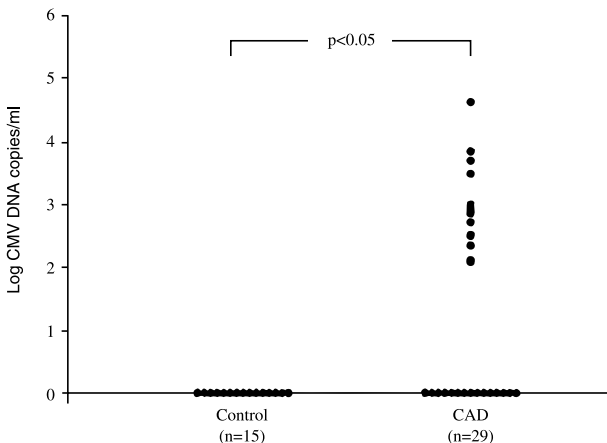
討論

我們的研究證實：1. 以抗體有無來顯示CMV 盛行率高低狀況與罹患CAD 與否無關。2. CMV PCR 陽性率在兩組間無顯著差異。3. CAD 組CMV 病毒量比對照組顯著增加。

一個從未暴露於CMV 存在的環境下之健康個體，是不會產生對抗CMV 抗體，故此抗體可作為感染CMV 的指標。體內產生的抗體主要又分成IgM 和IgG，分別代表首次感染或再復發以及二次感染或持續性感染。由於CAD 屬於慢性

表四：比較冠心病組與對照組CMV IgG、hs-CRP、IL-6之相關性

Group	Item			
	CMV IgG and IL-6	CMV IgG and hs-CRP	CMV IgG and PCR	IL-6 and hs-CRP
Control				
r value	0.249	0.045	0.044	0.119
p value	0.055	0.737	0.737	0.370
CAD				
r value	0.102	0.077	-0.128	0.634
p value	0.335	0.467	0.222	<0.001
Total				
r value	0.178	0.127	-0.047	0.503
p value	0.028	0.119	0.561	<0.001



圖一：CMV nested PCR 陽性個體血中CMV 病毒表現量

發炎性疾病，整個致病流程是個緩慢的過程，若CMV感染與CAD有關，應屬於長期感染CMV所誘發引起的疾病，與首次感染無關。故我們僅選取anti-CMV IgG抗體來代表個體的感染狀況。在不同族群裡，CMV的盛行率皆隨年齡增加而上升，然而在高度開發國家，盛行率較低，反之則較高⁵⁸⁻⁶¹。在台灣，CMV盛行率高達90%以上，且隨著年齡，盛行率有顯著增加的情形^{62, 63}。本次實驗數據對照組、CAD組的CMV抗體陽性率皆高達97%，符合前人論述，兩組間並無差異。然而若將抗體效價劃分四分區評估勝率比，則可發現CAD組在高抗體效價區的百分比有逐漸提高的趨勢（表三），這可能因為CAD持續著感染個體，造成CMV抗體效價偏高的緣故。雖然抗體的存在代表個體曾經感染過

CMV，但正常人多數是無症狀感染，CMV轉而潛伏在體內，伺機而動。為釐清這個問題，我們以nested PCR檢測個體血中CMV是否存在，然而陽性率分別只有25%（對照組）與31.2%（CAD組），未達統計學意義。雖然，研究個體大部分都感染過CMV，然而由PCR的結果看來，與CMV抗體表現並不一致，類似的結果亦出現在學者的研究中。可能因為CMV潛伏細胞種類太廣、或是PCR使用的標的基因敏感度不夠，或者是CMV潛伏感染時僅嵌入部分基因而導致DNA萃取過程中流失PCR標靶基因^{15, 64-67}。

IL-6、hs-CRP咸認為與CAD有關，因此我們亦比對了CMV抗體、CMV PCR結果在兩組間的表現，並進一步分析相關性。一如預期，IL-6與hs-CRP在CAD組呈現高度相關（ $r=0.634$, $p<0.001$ ）。另外，在對照組中，CMV抗體與IL-6相關性也有明顯增加（ $r=0.249$, $p=0.055$ ），且就總體而言，CMV抗體與IL-6相關性已有顯著的意義（ $r=0.178$, $p=0.028$ ）。這可能是因為CMV感染誘發宿主產生免疫反應的結果，然而hs-CRP由於是發炎反應的下游物質，因此相關性就不若IL-6來得顯著。

CMV感染與CAD疾病之間到底有無關連，相關研究各有數據佐證^{52, 68-72}。為釐清這個問題，本實驗除了顯示出抗體效價的差別外，亦將CMV在血中的病毒量納入衡量範圍。臨床上，用以定量CMV的方式不外乎三種，一為CMV

pp65 antigenemia，敏感度低、專一性高。二為 real time PCR，敏感度高（視標的基因種類而有不同的敏感度）、專一性較低。三為NASBA偵測法測pp67 mRNA，敏感度低、專一性較最高。本實驗採用標的基因為UL83之 real time PCR方式檢測CMV病毒量，UL83為pp65抗原的來源基因，因此與pp65 antigenemia test相關性相當高，敏感度或許不若nested PCR但是就專一性而言，絕對比nested PCR來的強。結果顯示：25個nested PCR (+) 對照組，偵測不到CMV病毒量。反之，29個nested PCR (+) CAD組，有14個可偵測到病毒量，範圍由數百至數萬間。這表示CAD組中，CMV病毒量較對照組顯著增加（圖一）。至於其他nested PCR (-) 檢體，則完全測不到病毒量（Data not shown）。

儘管CMV感染與CAD疾病的關係，紛紛擾擾、眾說紛紛。但是藉由病毒定量的方式，本實驗證實CAD患者體內帶有CMV病毒的量的確比對照組為高。這是因為CMV長期感染再加上其他危險因子而導致CAD疾病產生，還是CAD患者本身免疫力相對於對照組為低，而使潛伏性感染的CMV有可趁之機，再度活化感染宿主，因而造成CMV病毒量增加？若能釐清此問題，或許CMV在CAD疾病所扮演的角色就有柳暗花明的一天。

參考文獻

- Orlandi A, Bochaton-Piallat ML, Gabbiani G, Spagnoli LG. Aging, smooth muscle cells and vascular pathobiology: Implications for atherosclerosis. *Atherosclerosis* 2006; 188: 221-30.
- Rauscher FM, Goldschmidt-Clermont PJ, Davis BH, et al. Aging, progenitor cell exhaustion, and atherosclerosis. *Circulation* 2003; 108: 457-63.
- Robert L. Aging of the vascular-wall and atherosclerosis. *Exp Gerontol* 1999; 34: 491-501.
- Pessina AC. [Hypertension and atherosclerosis]. *Cardiologia* 1999; 44 Suppl 1: 259-63.
- Schroeder EB, Liao D, Chambless LE, Prineas RJ, Evans GW, Heiss G. Hypertension, blood pressure, and heart rate variability: the Atherosclerosis Risk in Communities (ARIC) study. *Hypertension* 2003; 42: 1106-11.
- Steinberger J, Daniels SR. Obesity, insulin resistance, diabetes, and cardiovascular risk in children: an American Heart Association scientific statement from the Atherosclerosis, Hypertension, and Obesity in the Young Committee (Council on Cardiovascular Disease in the Young) and the Diabetes Committee (Council on Nutrition, Physical Activity, and Metabolism). *Circulation* 2003; 107: 1448-53.
- Mori S, Saito Y. Cytokine and atherosclerosis: a possible role of osteopontin in development in diabetic macroangiopathy. *Nihon Rinsho Meneki Gakkai Kaishi* 2000; 23: 613-7.
- Liu Y, Berthier-Schaad Y, Fallin MD, et al. IL-6 haplotypes, inflammation, and risk for cardiovascular disease in a multiethnic dialysis cohort. *J Am Soc Nephrol* 2006; 17: 863-70.
- Piechota W, Piechota W. [Correlation of high-sensitivity CRP concentration with the extent of coronary atherosclerosis in men with symptoms of ischemic heart disease]. *Pol Merkuriusz Lek* 2005; 18: 511-5.
- Su TC, Jeng JS, Wang JD, et al. Homocysteine, circulating vascular cell adhesion molecule and carotid atherosclerosis in postmenopausal vegetarian women and omnivores. *Atherosclerosis* 2006; 184: 356-62.
- Guech-Ongey M, Brenner H, Twardella D, Hahmann H, Rothenbacher D. Role of cytomegalovirus sero-status in the development of secondary cardiovascular events in patients with coronary heart disease under special consideration of diabetes. *Int J Cardiol* 2006; 111: 98-103.
- Siscovick DS, Schwartz SM, Corey L, et al. Chlamydia pneumoniae, herpes simplex virus type 1, and cytomegalovirus and incident myocardial infarction and coronary heart disease death in older adults: the Cardiovascular Health Study. *Circulation* 2000; 102: 2335-40.
- Matthews-Greer JM, Eggerstedt JM, Gonzalez E, Jamison RM, Washington CD. Investigation of the prevalence of cardiovascular-associated cytomegalovirus among patients with coronary artery disease at Louisiana State University Medical Center at Shreveport. *J Infect Dis* 1998; 178: 1860-1.
- Sorlie PD, Nieto FJ, Adam E, Folsom AR, Shahar E, Massing M. A prospective study of cytomegalovirus, herpes simplex virus 1, and coronary heart disease: the atherosclerosis risk in communities (ARIC) study. *Arch Intern Med* 2000; 160: 2027-32.
- Slobodman B, Stern JL, Cunningham AL, Abendroth A, Abate DA, Mocarski ES. Impact of human cytomegalovirus latent infection on myeloid progenitor cell gene expression. *J Virol* 2004; 78: 4054-62.
- Kondo K, Kaneshima H, Mocarski ES. Human cytomegalovirus latent infection of granulocyte-macrophage progenitors. *Proc Natl Acad Sci USA* 1994; 91: 11879-83.
- Hayashi K, Saze K, Uchida Y. Studies of latent cytomegalovirus infection: the macrophage as a virus-harboring cell. *Microbiol Immunol* 1985; 29: 625-34.
- Jarvis MA, Nelson JA. Human cytomegalovirus persistence and latency in endothelial cells and macrophages. *Curr Opin Microbiol* 2002; 5: 403-7.
- Froberg MK, Adams A, Seacotte N, Parker-Thornburg J, Kolattukudy P. Cytomegalovirus infection accelerates inflam-

- mation in vascular tissue overexpressing monocyte chemoattractant protein-1. *Circ Res* 2001; 89: 1224-30.
20. Zhu J, Quyyumi AA, Norman JE, Csako G, Epstein SE. Cytomegalovirus in the pathogenesis of atherosclerosis: the role of inflammation as reflected by elevated C-reactive protein levels. *J Am Coll Cardiol* 1999; 34: 1738-43.
 21. Hunninghake GW, Monick MM, Geist LJ. Cytomegalovirus infection. Regulation of inflammation. *Am J Respir Cell Mol Biol* 1999; 21: 150-2.
 22. Horvath R, Cerny J, Benedik J, Jr., Hokl J, Jelinkova I, Benedik J. The possible role of human cytomegalovirus (HCMV) in the origin of atherosclerosis. *J Clin Virol* 2000; 16: 17-24.
 23. Lunardi C, Bason C, Corrocher R, Puccetti A. Induction of endothelial cell damage by hCMV molecular mimicry. *Trends Immunol* 2005; 26: 19-24.
 24. Froberg MK, Seacotte N, Dahlberg E. Cytomegalovirus seropositivity and serum total cholesterol levels in young patients. *Ann Clin Lab Sci* 2001; 31: 157-61.
 25. Carlquist JF, Muhlestein JB, Horne BD, et al. Cytomegalovirus stimulated mRNA accumulation and cell surface expression of the oxidized LDL scavenger receptor, CD36. *Atherosclerosis* 2004; 177: 53-9.
 26. Stoll G, Bendszus M. Inflammation and atherosclerosis: novel insights into plaque formation and destabilization. *Stroke* 2006; 37: 1923-32.
 27. Matsuura E, Kobayashi K, Hurley BL, Lopez LR. Atherogenic oxidized low-density lipoprotein/beta2-glycoprotein I (oxLDL/beta2GPI) complexes in patients with systemic lupus erythematosus and antiphospholipid syndrome. *Lupus* 2006; 15: 478-83.
 28. Matsuura E, Kobayashi K, Tabuchi M, Lopez LR. Oxidative modification of low-density lipoprotein and immune regulation of atherosclerosis. *Prog Lipid Res* 2006; 47: 2208-14.
 29. Yazici M, Demircan S, Ibrahimli F, Aksakal E, Sahin M, Sagkan O. [The importance of lipoprotein(a) in pathogenesis of the high risk unstable angina]. *Anadolu Kardiyol Derg* 2006; 6: 13-7.
 30. Levi M. CMV endothelitis as a factor in the pathogenesis of atherosclerosis. *Cardiovasc Res* 2001; 50: 432-3.
 31. Zhu J, Shearer GM, Norman JE, et al. Host response to cytomegalovirus infection as a determinant of susceptibility to coronary artery disease: sex-based differences in inflammation and type of immune response. *Circulation* 2000; 102: 2491-6.
 32. Zhu J, Shearer GM, Marincola FM, et al. Discordant cellular and humoral immune responses to cytomegalovirus infection in healthy blood donors: existence of a Th1-type dominant response. *Int Immunol* 2001; 13: 785-90.
 33. Harmenberg J, Brytting M. Limitations of cytomegalovirus testing. *Antimicrob Agents Chemother* 1999; 43: 1528-9.
 34. Sandin RL, Rodriguez ER, Rosenberg E, et al. Comparison of sensitivity for human cytomegalovirus of the polymerase chain reaction, traditional tube culture and shell vial assay by sequential dilutions of infected cell lines. *J Virol Methods* 1991; 32: 181-91.
 35. Stirk PR, Griffiths PD. Comparative sensitivity of three methods for the diagnosis of cytomegalovirus lung infection. *J Virol Methods* 1988; 20: 133-41.
 36. Degre M, Kristiansen KI, Rollag H, Holter E, Nordal KP. Detection of human cytomegalovirus (HCMV) pp67-mRNA and pp65 antigenemia in relation to development of clinical HCMV disease in renal transplant recipients. *Clin Microbiol Infect* 2001; 7: 254-60.
 37. Piiparinen H, Hockerstedt K, Gronhagen-Riska C, Lappalainen M, Suni J, Lautenschlager I. Comparison of plasma polymerase chain reaction and pp65-antigenemia assay in the quantification of cytomegalovirus in liver and kidney transplant patients. *J Clin Virol* 2001; 22: 111-6.
 38. Mazzulli T, Drew LW, Yen-Lieberman B, et al. Multicenter comparison of the digene hybrid capture CMV DNA assay (version 2.0), the pp65 antigenemia assay, and cell culture for detection of cytomegalovirus viremia. *J Clin Microbiol* 1999; 37: 958-63.
 39. Hernando S, Folgueira L, Lumberras C, et al. Comparison of cytomegalovirus viral load measure by real-time PCR with pp65 antigenemia for the diagnosis of cytomegalovirus disease in solid organ transplant patients. *Transplant Proc* 2005; 37: 4094-6.
 40. Herrmann B, Larsson VC, Rubin CJ, et al. Comparison of a duplex quantitative real-time PCR assay and the COBAS Amplicor CMV Monitor test for detection of cytomegalovirus. *J Clin Microbiol* 2004; 42: 1909-14.
 41. Boeckh M, Huang M, Ferrenberg J, et al. Optimization of quantitative detection of cytomegalovirus DNA in plasma by real-time PCR. *J Clin Microbiol* 2004; 42: 1142-8.
 42. Nitsche A, Oswald O, Steuer N, et al. Quantitative real-time PCR compared with pp65 antigen detection for cytomegalovirus (CMV) in 1122 blood specimens from 77 patients after allogeneic stem cell transplantation: which test better predicts CMV disease development? *Clin Chem* 2003; 49: 1683-5.
 43. Alice T, Enrietto M, Pittaluga F, et al. Quantitation of cytomegalovirus DNA by real-time polymerase chain reaction in peripheral blood specimens of patients with solid organ transplants: comparison with end-point PCR and pp65 antigen test. *J Med Virol* 2006; 78: 915-22.
 44. Yun Z, Lewensohn-Fuchs I, Ljungman P, Vahlne A. Real-time monitoring of cytomegalovirus infections after stem cell transplantation using the TaqMan polymerase chain reaction assays. *Transplantation* 2000; 69: 1733-6.
 45. Taylor-Wiedeman J, Sissons JG, Borysiewicz LK, Sinclair JH. Monocytes are a major site of persistence of human cytomegalovirus in peripheral blood mononuclear cells. *J Gen Virol* 1991; 72: 2059-64.
 46. Stanier P, Taylor DL, Kitchen AD, Wales N, Tryhorn Y, Tys AS. Persistence of cytomegalovirus in mononuclear cells in peripheral blood from blood donors. *Bmj* 1989; 299: 897-8.
 47. Bevan IS, Daw RA, Day PJ, Ala FA, Walker MR. Polymerase chain reaction for detection of human cytomegalovirus infection in a blood donor population. *Br J Haematol* 1991; 78(1): 94-9.

48. Larsson S, Soderberg-Naucleer C, Wang FZ, Moller E. Cytomegalovirus DNA can be detected in peripheral blood mononuclear cells from all seropositive and most seronegative healthy blood donors over time. *Transfusion* 1998; 38: 271-8.
49. Bitsch A, Kirchner H, Dupke R, Bein G. Failure to detect human cytomegalovirus DNA in peripheral blood leukocytes of healthy blood donors by the polymerase chain reaction. *Transfusion* 1992; 32: 612-7.
50. Urushibara N, Kwon KW, Takahashi TA, Sekiguchi S. Human cytomegalovirus DNA is not detectable with nested double polymerase chain reaction in healthy blood donors. *Vox Sang* 1995; 68: 9-14.
51. Bevan IS, Walker MR, Daw RA. Detection of human cytomegalovirus DNA in peripheral blood leukocytes by the polymerase chain reaction. *Transfusion* 1993; 33: 783-4.
52. Schlitt A, Blankenberg S, Weise K, et al. Herpesvirus DNA (Epstein-Barr virus, herpes simplex virus, cytomegalovirus) in circulating monocytes of patients with coronary artery disease. *Acta Cardiol* 2005; 60: 605-10.
53. Roback JD, Drew WL, Laycock ME, Todd D, Hillyer CD, Busch MP. CMV DNA is rarely detected in healthy blood donors using validated PCR assays. *Transfusion* 2003; 43: 314-21.
54. Gault E, Michel Y, Dehee A, Belabani C, Nicolas JC, Garbarg-Chenon A. Quantification of human cytomegalovirus DNA by real-time PCR. *J Clin Microbiol* 2001; 39: 772-5.
55. Tanaka N, Kimura H, Iida K, et al. Quantitative analysis of cytomegalovirus load using a real-time PCR assay. *J Med Virol* 2000; 60: 455-62.
56. Greenlee DJ, Fan H, Lawless K, Harrison CR, Gulley ML. Quantitation of CMV by real-time PCR in transfusable RBC units. *Transfusion* 2002; 42: 403-8.
57. Demmler GJ, Buffone GJ, Schimbor CM, May RA. Detection of cytomegalovirus in urine from newborns by using polymerase chain reaction DNA amplification. *J Infect Dis* 1988; 158: 1177-84.
58. Chauhan M, Barratt CL, Cooke S, Cooke ID. Screening for cytomegalovirus antibody in a donor insemination program: difficulties in implementing The American Fertility Society guidelines. *Fertil Steril* 1989; 51: 901-2.
59. Nicolle LE, Minuk GY, Postl B, Ling N, Madden DL, Hoofnagle JH. Cross-sectional seroepidemiologic study of the prevalence of cytomegalovirus and herpes simplex virus infection in a Canadian Inuit (Eskimo) community. *Scand J Infect Dis* 1986; 18: 19-23.
60. Tantivanich S, Suphadtanaphongs V, Siripanth C, et al. Prevalence of cytomegalovirus antibodies among various age groups of Thai population. *Southeast Asian J Trop Med Public Health* 1999; 30: 265-8.
61. Emery VC. Cytomegalovirus and the aging population. *Drugs Aging* 2001; 18: 927-33.
62. Lu SC, Chin LT, Wu FM, et al. Seroprevalence of CMV antibodies in a blood donor population and premature neonates in the south-central Taiwan. *Kaohsiung J Med Sci* 1999; 15: 603-10.
63. Shen CY, Chang WW, Chang SF, Chao MF, Huang ES, Wu CW. Seroepidemiology of cytomegalovirus infection among children between the ages of 4 and 12 years in Taiwan. *J Med Virol* 1992; 37: 72-5.
64. Sinclair J, Sissons P. Latency and reactivation of human cytomegalovirus. *J Gen Virol* 2006; 87: 1763-79.
65. Reeves MB, MacAry PA, Lehner PJ, Sissons JG, Sinclair JH. Latency, chromatin remodeling, and reactivation of human cytomegalovirus in the dendritic cells of healthy carriers. *Proc Natl Acad Sci U S A*. 2005; 102(11): 4140-5.
66. Froberg MK. Review: CMV escapes! *Ann Clin Lab Sci* 2004; 34: 123-30.
67. Sissons JG, Bain M, Wills MR. Latency and reactivation of human cytomegalovirus. *J Infect* 2002; 44: 73-7.
68. Eryol NK, Kilic H, Gul A, et al. Are the high levels of cytomegalovirus antibodies a determinant in the development of coronary artery disease? *Int Heart J* 2005; 46: 205-9.
69. Rahel BM, Visseren FL, Suttorp MJ, et al. Cytomegalovirus and Chlamydia pneumoniae as predictors for adverse events and angina pectoris after percutaneous coronary intervention. *Am Heart J* 2004; 148: 670-5.
70. Muhlestein JB, Anderson JL. Chronic infection and coronary artery disease. *Cardiol Clin* 2003; 21: 333-62.
71. Rothenbacher D, Brenner H, Hoffmeister A, Mertens T, Persson K, Koenig W. Relationship between infectious burden, systemic inflammatory response, and risk of stable coronary artery disease: role of confounding and reference group. *Atherosclerosis* 2003; 170: 339-45.
72. Khairy P, Rinfret S, Tardif JC, et al. Absence of association between infectious agents and endothelial function in healthy young men. *Circulation* 2003; 107: 1966-71.

The Correlation Between Cytomegalovirus Infection and Coronary Artery Disease

Yung-Liang Liao¹, Yi-Chueh Yang¹, Ting-Chun Hung¹, Hui-Yi Yap²,
Chia-Fong Chen³, Tung-Nan Liao³, and Ching-Nan Lin¹

¹*Department of Pathology, Chi-Mei Foundation Hospital, Tainan, Taiwan,*

²*Department of Internal Medicine, Chi-Mei Foundation Hospital.*

³*Department of Medical Technology, Chung-Hwa College of Medical Technology*

Primary CMV infection is usually asymptomatic and results in latent, lifelong persistence of the viral genome. Periodically, the virus might reactivate from latency and regain its ability to multiply. CMV is a major cause of life-threatening illness in immunocompromised patients. Various lines of evidence suggest that chronic inflammation is involved in the initiation and progression of atherosclerosis and its complications including coronary artery disease (CAD). Several infectious agents, particularly CMV, have been suggested to play a role in the development of atherosclerosis. In this study, we measured serum levels of IL-6, anti-CMV IgG (α CMV IgG), high sensitivity C-reactive protein (hs-CRP) by ELISA or nephelometry, CMV qualitative analysis by nested PCR, and CMV viral load quantitative analysis by real time PCR in 93 CAD patients and 60 normal healthy controls. Our study showed that the levels of IL-6, and hs-CRP were significantly increased in patients with CAD ($p=0.002$ and $p<0.001$, respectively). There was no statistical significance in α CMV IgG positive rate in both groups, however, there was a trend for higher antibody titers in the CAD group according to the quartile levels of analysis ($p=0.078$). In nested PCR positive cases, the CMV viral loads were significantly elevated in CAD patients than in normal controls ($p<0.05$); meanwhile, there was no difference in CMV positive rates between the two groups using nested PCR qualitative assay. Conclusion: In CMV nested PCR positive cases, CMV viral loads were significantly elevated in CAD patients than in normal controls. However, there were no differences in CMV nested PCR and α CMV IgG positive rates between the two groups. (J Intern Med Taiwan 2007; 18: 88-96)