

***Cryptococcus gattii* Meningitis Developed after Pneumonectomy in A Case of Necrotizing Pneumonia**

Chih-Chen Lin^{1*}, Hsiang-Kuang Tseng^{1,2,3*}, Wei-Sheng Wang^{1,2,3}, Yee-Chun Chen⁴,
Tseng-Yu Huang^{1,2,3}, Alice Ying-Jung Wu^{1,2,3}, and Chang-Pan Liu^{1,2,3}

¹Section of Infectious Diseases, Department of Internal Medicine, Mackay Memorial Hospital;

²Department of Medicine, Mackay Medical College;

³Mackay Junior College of Medicine, Nursing, and Management;

⁴Division of Infectious Diseases, Department of Internal Medicine, National Taiwan University Hospital and College of Medicine

Abstract

Cryptococcus gattii infection has rarely been reported in Taiwan. We report a case of 32-year-old immunocompetent male Vietnamese referred for pneumonectomy under the impression of necrotizing pneumonia. Pleural effusion was collected during the pneumonectomy and the culture of pleural effusion yielded *Cryptococcus*. Although oral fluconazole 200 mg daily was prescribed, meningitis developed 10 days after pneumonectomy. Culture of cerebrospinal fluid (CSF) yielded *Cryptococcus*. The patient received amphotericin B plus flucytosine for two weeks as antifungal induction therapy, followed by intravenous fluconazole 400 mg per day as consolidation therapy. The molecular typing of *Cryptococcus* was *C. gattii* VGII. He was discharged on the 40th day after admission and was prescribed oral fluconazole 200 mg daily for 110 days at outpatient department. No neurological sequela was found at the time of last follow-up. Importantly, *Cryptococcus* has tendency to infect the central nervous system (CNS), especially the subspecies *C. gattii*, which could be differentiated from *C. neoformans* by its activity in the medium containing canavanine, glycine and bromothymol blue (CGB). Patients with necrotizing pneumonia caused by *C. gattii* should be treated as central nervous system infection. In conclusion, appropriate fungicidal agents to cover CNS infection should be administered for this kind of patient from the time of first disease onset. (J Intern Med Taiwan 2014; 25: 30-35)

Key Words: *Cryptococcus*, *Cryptococcus gattii*, Meningitis, Necrotizing pneumonia

Introduction

Cryptococcus infection is life threatening and is related to severe lung and central nervous system complications. It also has unique microbiological, epidemiological, clinical presentations and

outcomes¹. Among members of *Cryptococcus neoformans-Cryptococcus gattii* species complex, *C. neoformans* is distributed worldwide whereas *C. gattii* is considered to be more prevalent in the subtropical and tropical areas including Taiwan. The two species of *Cryptococcus* can be differenti-

Reprint requests and correspondence : Dr. Chang-Pan Liu

Address : Section of Infectious Diseases, Department of Internal Medicine, Mackay Memorial Hospital, No. 92, Sec. 2, Zhongshan N. Rd., Taipei City 10449, Taiwan

*Chih-Chen Lin and Hsiang-Kuang Tseng are equal contribution.

ated using a solid agar *medium* containing canavanine, glycine and bromothymol blue (CGB)². Furthermore, by using orotidine monophosphate pyrophosphorylase (URA5) gene restriction fragment length polymorphism analysis, they can be further divided into eight genotypes, including four types of *C. neoformans* (VNI, VNII, VNIII, and VNIV) and four types of *C. gattii* (VGI, VGII, VGIII, and VGIV)³. *C. gattii* VGI molecular type is traditionally considered to be the most prevalent, although *C. gattii* VGII molecular type caused the Vancouver Island outbreak^{4,5}. The VGIII and VGIV molecular types of *C. gattii* were commonly isolated from HIV-infected patients in Africa and Northern America^{6,7}.

C. gattii was isolated more often than *C. neoformans* in immunocompetent patients⁶. Patients infected with *C. gattii* were younger, more likely to have no underlying conditions and more likely to have meningoencephalitis¹. In common model, rats infected with the *C. gattii* infections result in more cryptococcoma complicated with hemorrhage

and neurological sequelae requiring longer treatment duration⁷⁻⁹. Whether host immune function or different subtypes of *C. gattii* contribute more to poor prognosis is still in debate⁵⁻⁷.

In 1990, *C. gattii* was isolated from *Eucalyptus camaldulensis*; subsequently, it was believed that *C. gattii* spread through the exportation and flowering of *Eucalyptus* species^{10,11}. An environmental study in 2007 also isolated *C. gattii* from a broad range of trees in Canada¹². This implied that plants are probably the niche of *C. gattii*. In 1999, *C. gattii* caused an outbreak in Vancouver Island (British Columbia), Canada. Since then, numerous studies have revealed more information about *C. gattii*; which challenged what we knew about *C. gattii* in the past^{4,6}. For example, genealogy and phylogenetic analyses revealed that *C. gattii* VGII found in the temperate regions of British Columbia most likely emerged from the tropical Amazon rainforest¹³.

We introduce a 32 year-old Vietnamese lumberman who had necrotizing pneumonia related to *Cryptococcus gattii* VGII. It was interesting that

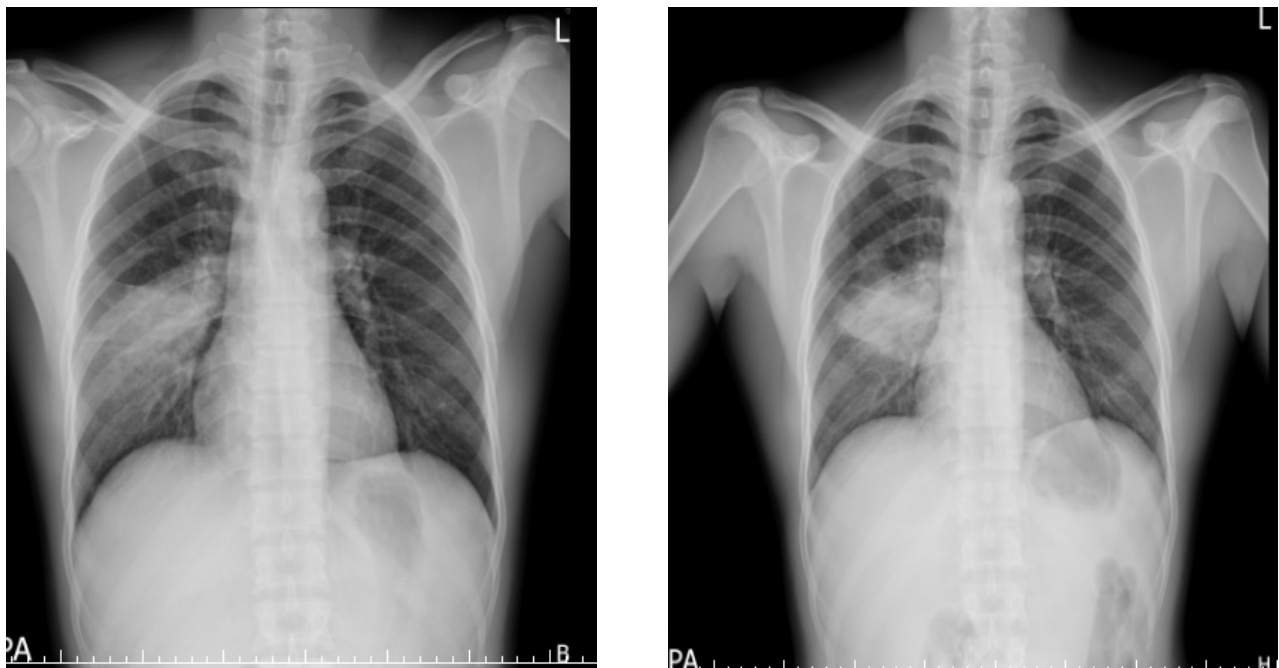


Figure 1. (A) The chest X-ray 2 weeks before referral presents right lower lobe opacity; (B) The chest X-ray on referral day presents stationary right lower lobe lesion compared with previous film.

his meningitis developed 10 days after pneumonec-
tomy. Even though the diagnosis was made late and
the surgeon did not use fungicidal agents initially,
the patient recovered without neurologic sequela.
Because *C. gattii* present with unique characteris-
tics and clinical manifestations, it is important to
discriminate it from *C. neoformans*.

Case report

A 32 year-old male Vietnamese came to
Taitung, Taiwan and worked as a lumberman for
five years. He suffered from intermittent fever
and persistent cough with yellow sputum for two
weeks. He had generalized malaise and poor appe-
tite. He denied body weight loss, chest tightness,
gastrointestinal symptoms or genitourinary symp-
toms. Then he went to clinic for further evaluation.
The chest radiograph showed right lower lobe
opacity (Figure 1A). Oral amoxicillin/clavunilate
and roxythromycin were empirically prescribed.
Initially neither bacteria nor mycobacterium was
found in the sputum Gram stain and acid fast stain.
Fever subsided but he had sustained productive
cough. After two weeks of oral antibiotics treat-
ment, infiltrates persisted on the chest radiograph
(Figure 1B) and clinical symptoms progressed.
Then he was admitted for assessment. He had
history of *Blastocystis hominis* infection which
was diagnosed through immigrant health examina-
tion, for which he received complete treatment with
metronidazole. He smoked about one pack per day
for 15 years. The blood examination showed white
blood cell count 12,400 with 81.9% neutrophils but
the rest of the data were in normal range. Contrast-
enhanced computed tomography (CT) of the chest
revealed heterogeneous lesions on superior segment
of the right lower lung, suspicious of necrotizing
pneumonia (Figure 2). Empiric piperacillin/tazo-
bactom and levofloxacin were prescribed. Bron-
choscopy showed a protruding hypervascular
lesion situated at RB 6 orifice (Figure 3), which is

the superior segment of the right lower lobe. The
sputum cultures for bacteria and *Mycobacterium
tuberculosis* yielded no growth and cytology showed
no malignant cell. Under the tentative diagnosis
of necrotizing pneumonia which failed medical
treatment, he was referred to a medical center for
surgical intervention. Pneumonec-
tomy was done on the 4th day after referral. Pathology of the lung
tissue showed cryptococcosis with mucin pools
surrounded by prominent inflammation. Culture



Figure 2. The computed tomography presents a hetero-
geneous lesion in superior segment of right
lower lobe, suspected to be necrotizing pneu-
monitis.

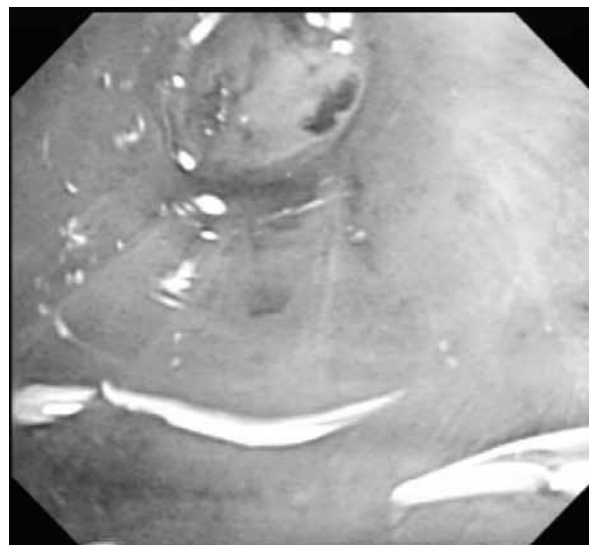


Figure 3. The bronchoscope shows protruding mass
with hypervascular lesion on RB 6 orifice.

of the pleural effusion showed *Cryptococcus*. Oral fluconazole 200 mg daily was prescribed. However, fever flared up along with headache, nausea and vomiting on the 10th day after surgery. Brain CT showed no specific finding. Meningitis was suspected and lumbar puncture was done. Cerebrospinal fluid (CSF) study revealed protein 53 mg/dL (10~45 mg/dL), glucose 62 mg/dL (45~75 mg/dL), WBC 84/ml with lymphocyte 66/ml and neutrophil 14/ml. India ink was negative but the *Cryptococcus* antigen titer in both CSF and serum were positive and were 1:8 and 1:64 respectively. Amphotericin B and flucytosine were prescribed as antifungal induction therapy. Both pleural effusion and CSF culture yielded *Cryptococcus* species. Because the yeast turned CGB medium blue, it was identified as *Cryptococcus gattii*. Molecular typing with URA 5 identified the species as VGII. Antifungal susceptibility obtained by a commercially prepared, dried colorimetric microdilution panel (Sensititre YeastOne, Thermo-Fisher Scientific, West Sussex, UK) was shown in Table 1. No yeast was identified in the blood culture bottle. After two weeks of induction therapy with amphotericin B 50 mg daily and flucytosine 2 gm every six hours, the patient felt better. We switched to fluconazole 400 mg daily as consolidation therapy. He was discharged on the 40th day after referral. He received oral fluconazole 200 mg daily in the outpatient clinic of a local hospital in Taitung. No neurological sequela was found after a follow-up period of 110 days.

Discussion

Both Vietnam and Taitung, Taiwan are in the tropics, with climate suitable for the growth of *C. gattii*^{1,14}. This Vietnamese patient has been working in Taiwan for 5 years as a lumberman. He was in the risk of exposure to certain kinds of plants which might possess *C. gattii* although there is a lack of environmental survey studies in Taiwan. Since articles report the delayed onset *C. gattii*

Table 1. Antifungal susceptibility of the pleural effusion and CSF *Cryptococcus gattii* isolates by a commercial microdilution panel Sensititre YeastOne

| | Pleural effusion | CSF |
|----------------|------------------|-------------|
| Antifungal | MIC (µg/mL) | MIC (µg/mL) |
| Amphotericin B | 0.25 | 0.25 |
| Flucytosine | 1 | 1 |
| Fluconazole | 16 | 16 |
| Voriconazole | 0.12 | 0.12 |

infection which occurred more than 10 months after traveling to *C. gattii* endemic area^{15,16}, it is unknown whether the patient acquired *C. gattii* from Taiwan or Vietnam. Further genetic analysis such as multi-locus sequence typing to confirm the original source of pathogen was needed.

It is an unusual for cryptococcal meningitis to develop after pneumonectomy. Several possibilities could have contributed to the patient's meningitis, including fungal load, molecular types, lack of initial application of fungicidal agent, and transient fungemia post pneumonectomy. A rat model was built to explore the pathogenesis of *C. gattii* infection⁸. Rats developed gross lung lesions with dissemination to brain, kidney and spleen only when the yeast inoculated was more than 10⁷ colony forming units (CFUs) per 0.1 ml. Different clinical manifestation and prognosis among *C. gattii* correlated with different subtype and mating type of *Cryptococcus*. The molecular type VGII (Colombia VGIIa- MAT *alpha* and VGIIb- MAT *alpha*) has tendency to induce late onset CNS infection. In addition, the lack of fungicidal agents use initially also contributed to the disseminated infection in our case. According to the 2010 IDSA guideline for cryptococcal infection, severe pulmonary cryptococcal infection should be treated as CNS infection¹⁷. The IDSA guideline suggested two steps therapy. Induction therapy with amphotericin B 0.5~0.8 mg/kg/day intravenous infusion and flucytosine 25mg/

kg oral every six hours is suggested until patients become afebrile and the culture turn negative. Further antifungal consolidation therapy with oral fluconazole 400 mg daily is recommended for 8~10 weeks. Chen et al studied 86 patients who suffered from *C. gattii* infection in the lung or in the central nervous system¹⁸. In comparison with *C. neoformans*, it was found that infection with *C. gattii* required longer treatment duration; additionally, antifungal induction therapy with amphotericin B plus flucytosine led to better outcome in comparison to other antifungal agents¹⁸.

The Clinical and Laboratory Standards Institute (CLSI) does not provide clinical breakpoints (CBPs) for *Cryptococcus* species¹⁹; the epidemiologic cutoff values (ECVs) (highest wild type susceptibility endpoint) of antifungal susceptibility for reference was also not available for VGII^{20,21}. ECVs is the minimum inhibition concentration (MIC) values that captured >95% of the observed population in RPMI medium provided in recent studies^{20,21}. While CBPs predict the clinical outcome of therapy, the ECVs could monitor the emergence of strains with reduced susceptibility to the agent being evaluated due to mutation.

Finally, pneumonectomy might have resulted in transient fungemia which subsequently led to meningitis. In a murine inhalation model, *C. gattii* could be isolated from the brain despite consistent failure to recover infected cells from the blood.⁷ The authors speculate that transient fungemia may be the reason of the dissemination. Several factors may explain this phenomenon. First, *C. gattii* has ability to suppress the release of inflammatory factors⁷. Second, *C. gattii* could accommodate into macrophages, which allows it to cross the blood-brain barrier (Trojan's horse mechanism)^{7,22,23}. Hence, despite the low fungal load in blood, *C. gattii* could still effectively cross the BBB and cause CNS infection.

In conclusion, *C. gattii* infection is rare in Taiwan. Early diagnosis and prompt treatment are

important in CNS *C. gattii* infection. This case reminds us that severe pulmonary cryptococcal infection should be treated as CNS infection. The case also demonstrates that it is important to discriminate *C. gattii* from *C. neoformans* by CGB medium in clinical situations.

References

1. Tseng HK, Liu CP, Ho MW, et al. Microbiological, epidemiological, and clinical characteristics and outcomes of patients with cryptococcosis in Taiwan, 1997-2010. *PLoS One* 2013; 8: e61921.
2. Kwon-Chung KJ, Polacheck I, Bennett JE . Improved diagnostic medium for separation of *Cryptococcus neoformans* var. *neoformans* (serotypes A and D) and *Cryptococcus neoformans* var. *gattii* (serotypes B and C). *J Clin Microbiol* 1982; 15: 535-7.
3. Meyer W, Castaneda A, Jackson S, et al. Molecular typing of IberoAmerican *Cryptococcus neoformans* isolates. *Emerg Infect Dis* 2003; 9: 189-95.
4. Bartlett KH, Cheng PY, Duncan C, et al. A decade of experience: *Cryptococcus gattii* in British Columbia. *Mycopathologia* 2012; 173: 311-9.
5. Chambers C, MacDougall L, Li M, et al. Tourism and specific risk areas for *Cryptococcus gattii*, Vancouver Island, Canada. *Emerg Infect Dis* 2008; 14: 1781-3.
6. Harris J, Lockhart S, Chiller T . *Cryptococcus gattii*: where do we go from here? *Med Mycol* 2012; 50: 113-29.
7. Ngamskulrunroj P, Chang Y, Sionov E, et al. The primary target organ of *Cryptococcus gattii* is different from that of *Cryptococcus neoformans* in a murine model. *MBio* 2012; 3: e00103-12
8. Krockenberger MB, Malik R, Ngamskulrunroj P, et al. Pathogenesis of pulmonary *Cryptococcus gattii* infection: a rat model. *Mycopathologia* 2010; 170: 315-30.
9. Thompson GR, 3rd, Wiederhold NP, Najvar LK, et al. A murine model of *Cryptococcus gattii* meningoencephalitis. *J Antimicrob Chemother* 2012; 67: 1432-8.
10. Hagen F, Boekhout T . The search for the natural habitat of *Cryptococcus gattii*. *Mycopathologia* 2010; 170: 209-11.
11. Springer DJ, Chaturvedi V . Projecting global occurrence of *Cryptococcus gattii*. *Emerg Infect Dis* 2010; 16: 14-20.
12. Kidd SE, Chow Y, Mak S, et al. Characterization of environmental sources of the human and animal pathogen *Cryptococcus gattii* in British Columbia, Canada, and the Pacific Northwest of the United States. *Appl Environ Microbiol* 2007; 73: 1433-43.
13. Hagen F, Ceresini PC, Polacheck I, et al. Ancient Dispersal of the Human Fungal Pathogen *Cryptococcus gattii* from the Amazon Rainforest. *PLoS One* 2013; 8: e71148.
14. Chau TT, Mai NH, Phu NH, et al. A prospective descriptive study of cryptococcal meningitis in HIV uninfected patients in Vietnam - high prevalence of *Cryptococcus neoformans* var *grubii* in the absence of underlying disease. *BMC Infect*

- Dis 2010; 10: 199.
15. Georgi A, Schneemann M, Tintelnot K, et al. Cryptococcus gattii meningoencephalitis in an immunocompetent person 13 months after exposure. Infection 2009; 37: 370-3.
 16. Levy R, Pitout J, Long P, et al. Late presentation of Cryptococcus gattii meningitis in a traveller to Vancouver Island: A case report. Can J Infect Dis Med Microbiol 2007; 18: 197-9.
 17. Perfect JR, Dismukes WE, Dromer F, et al. Clinical practice guidelines for the management of cryptococcal disease: 2010 update by the infectious diseases society of america. Clin Infect Dis 2010; 50: 291-322.
 18. Chen SCA, Korman TM, Slavin MA, et al. Antifungal Therapy and Management of Complications of Cryptococcosis due to Cryptococcus gattii. Clin Infect Dis 2013; 57: 543-51.
 19. Clinical and Laboratory Standards Institute (CLSI). Reference method for broth dilution antifungal susceptibility testing of yeasts; approved standard. 3rd ed. Wayne, PA, Clinical and Laboratory Standards Institute. 2008.
 20. Espinel-Ingroff A, Chowdhary A, Cuenca-Estrella M, et al. Cryptococcus neoformans-Cryptococcus gattii species complex: an international study of wild-type susceptibility endpoint distributions and epidemiological cutoff values for amphotericin B and flucytosine. Antimicrob Agents Chemother 2012; 56: 3107-13.
 21. Espinel-Ingroff A, Aller AI, Canton E, et al. Cryptococcus neoformans-Cryptococcus gattii species complex: an international study of wild-type susceptibility endpoint distributions and epidemiological cutoff values for fluconazole, itraconazole, posaconazole, and voriconazole. Antimicrob Agents Chemother 2012; 56: 5898-906.
 22. Charlier C, Nielsen K, Daou S, et al. Evidence of a role for monocytes in dissemination and brain invasion by Cryptococcus neoformans. Infect Immun 2009; 77: 120-7.
 23. Tseng HK, Huang SH, Shackleford G, et al. The blood-brain barrier: Targeting in the pathogenesis of C. neoformans meningitis and drug discovery. Anti-Infective Agents 2013; 11: 1-10.

格特隱球菌腦膜炎發生於壞死性肺炎肺切除術後

林志錚^{1*} 曾祥洸^{1,2*} 王威勝^{1,2} 陳宜君⁴ 黃增裕¹ 巫映蓉¹ 劉昌邦^{1,2,3}

¹馬偕紀念醫院 感染科

²馬偕醫學院醫學系

³馬偕護理專科學校

⁴台大醫院 感染科

摘要

在新型隱球菌及格特隱球菌複合種群 (*Cryptococcus neoformans*-*Cryptococcus gattii* species complex) 之中，新型隱球菌的分布是全球性的，而格特隱球菌的分布主要被認為是在熱帶及亞熱帶地區包括台灣。格特隱球菌 (*Cryptococcus gattii*) 的感染在台灣很少見，我們報告一個 32 歲免疫正常的越南男性因為壞死性肺炎轉至我們醫院進行肺切除術。肋膜積水培養出格特隱球菌。起初我們每日投與口服 fluconazole 200mg。病人於肺切除術後 10 日發生腦膜炎。腦脊髓液亦培養出之格特隱球菌與肋膜積水相同。病人接受 amphotericin B 和 flucytosine 治療兩週後，改成針劑 fluconazole 400mg 繼續治療。最終病人於住院後 40 天順利出院。經 110 天每日口服 fluconazole 200mg 後，門診追蹤發現無任何神經併發症之報告。格特隱球菌容易產生中樞神經之感染。因此嚴重的肺部隱球菌感染應當作中樞神經感染治療。

(* 林志錚、曾祥洸為共同第一作者。)